



This is a repository copy of *Production of alkenes and novel secondary products by P450 OleT JE using novel H<sub>2</sub>O<sub>2</sub>-generating fusion protein systems.*

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/112349/>

Version: Accepted Version

---

**Article:**

Matthews, S., Tee, K.L., Rattray, N.J. et al. (5 more authors) (2017) Production of alkenes and novel secondary products by P450 OleT JE using novel H<sub>2</sub>O<sub>2</sub>-generating fusion protein systems. FEBS Letters. ISSN 0014-5793

<https://doi.org/10.1002/1873-3468.12581>

---

**Reuse**

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

Received Date : 08-Nov-2016

Revised Date : 23-Jan-2017

Accepted Date : 26-Jan-2017

Article type : Research Letter

## Production of alkenes and novel secondary products by P450 OleT<sub>JE</sub> using novel H<sub>2</sub>O<sub>2</sub>-generating fusion protein systems

Sarah Matthews<sup>1</sup>, Kang Lan Tee<sup>1</sup>, Nicholas J. Rattray<sup>1</sup>, Kirsty J. McLean<sup>1</sup>, David Leys<sup>1</sup>, David A. Parker<sup>2</sup>, Richard T. Blankley<sup>3</sup> and Andrew W. Munro<sup>1\*</sup>

<sup>1</sup>Manchester Institute of Biotechnology, Centre for Synthetic Biology of Fine and Specialty Chemicals (SYNBIOCHEM), School of Chemistry, The University of Manchester, Manchester M1 7DN, United Kingdom. <sup>2</sup>The Westhollow Technology Center, Houston, Texas 77028-3101, USA. <sup>3</sup>Agilent Technologies UK Ltd., Lakeside, Cheadle Royal Business Park, Stockport, Cheshire SK8 3GR, United Kingdom.

\*Author for correspondence: Professor Andrew W. Munro, Phone 0044-161-3065151, Email: Andrew.Munro@Manchester.ac.uk

### Abstract

*Jeotgalicoccus* sp. 8456 OleT<sub>JE</sub> (CYP152L1) is a fatty acid decarboxylase cytochrome P450 that uses hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) to catalyse production of terminal alkenes, which are industrially important chemicals with biofuel applications. We report enzyme fusion systems in which *Streptomyces coelicolor* alditol oxidase (AldO) is linked to OleT<sub>JE</sub>. AldO oxidizes polyols (including glycerol), generating H<sub>2</sub>O<sub>2</sub> as a co-product and facilitating its use for efficient OleT<sub>JE</sub>-dependent fatty acid decarboxylation. AldO activity is regulatable by polyol substrate titration, enabling control over H<sub>2</sub>O<sub>2</sub> supply to minimise oxidative inactivation of OleT<sub>JE</sub> and prolong activity for increased alkene production. We also use these fusion systems to generate novel products from secondary turnover of 2-OH and 3-OH myristic acid primary products, expanding the catalytic repertoire of OleT<sub>JE</sub>.

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/1873-3468.12581

This article is protected by copyright. All rights reserved.

## Keywords

OleT<sub>JE</sub>, CYP152L1, cytochrome P450, alkene production, hydrogen peroxide, fusion enzymes

OleT<sub>JE</sub> peroxygenase P450, alkene production, fusion enzymes

## Abbreviations

BSTFA - N,O-bis(trimethylsilyl)trifluoroacetamide; DCM - dichloromethane; EDTA - ethylenediaminetetraacetic acid; IPTG - isopropyl 1-thio-β-D-galactopyranoside; OleT<sub>JE</sub> – cytochrome P450 OleT<sub>JE</sub> (CYP125L1) from *Jeotgalicoccus* sp. 8456; P450 – cytochrome P450; TMCS – trimethylchlorosilane; ΔALA – δ-aminolevulinic acid

## Introduction

The *Jeotgalicoccus* sp. 8456 OleT<sub>JE</sub> (CYP152L1) is a member of the cytochrome P450 (P450 or CYP) enzyme superfamily. P450s bind heme *b*, which is proximally coordinated by a cysteine thiolate and typically has a water ligand in the 6<sup>th</sup> position in the resting state (1). Substrate binding generally displaces the distal water ligand, converting low-spin (LS) ferric heme iron to the high-spin (HS) state (2). The HS heme has a more positive potential, facilitating heme iron reduction by a redox partner, binding of dioxygen to ferrous iron, and subsequent reduction and protonation steps resulting in formation of the ferryl-oxo porphyrin radical species compound I, which is the major substrate oxidizing species in P450s. P450s perform numerous oxidative (and other) reactions, including hydroxylation, demethylation, epoxidation and dehydrogenation of a vast range of substrates (3).

However, not all P450s are redox partner-dependent. Peroxygenase P450s function efficiently using only hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) to drive catalysis. Early peroxygenase studies focused on P450-BS<sub>β</sub> (CYP152A1) from *Bacillus subtilis* and P450-SP<sub>α</sub> (CYP152B1) from *Sphingomonas paucimobilis*. Initial studies suggested these enzymes were fatty acid hydroxylases, using the “peroxide shunt” mechanism to convert substrate-bound, ferric P450 directly to its ferric-hydroperoxo (compound 0) state, which then undergoes rapid protonation and dehydration to form compound I (4,5) (**Figure 1A**). However, Rude et al. subsequently demonstrated that P450-BS<sub>β</sub> also produced n-1 terminal alkenes alongside 2-OH and 3-OH fatty acids, suggesting that other CYP152 peroxygenases might also produce alkenes through oxidative decarboxylation of fatty acids (6). The *Jeotgalicoccus* OleT<sub>JE</sub> P450 was identified as a more efficient alkene producer, catalysing oxidative decarboxylation of fatty acids from ~C10-C20, generating terminal alkenes as major products, with smaller amounts of 2-OH and 3-OH fatty acids (6,7). Terminal alkenes have industrial uses in plastics, lubricants and as feedstocks for other compounds, e.g. alcohols and surfactants (8). Alkenes are also potential fuels, making OleT<sub>JE</sub> an attractive candidate for production of “drop-in” biofuels (9).

The structure and catalytic mechanism of OleT<sub>JE</sub> have been characterized (10-12), revealing how the fatty acid carboxylate binds close to the P450 heme (**Figure 1B**), as well as the different mechanisms by which OleT<sub>JE</sub> produces alkenes and hydroxylated fatty acids. OleT<sub>JE</sub> was reported to catalyse 97% conversion of 200 μM myristic acid (C14:0) in the presence of 500 μM H<sub>2</sub>O<sub>2</sub> (7). However, on an industrial scale excess H<sub>2</sub>O<sub>2</sub> may inhibit enzymatic activity (13). This phenomenon was demonstrated

in the *Clostridium acetobutylicum* peroxygenase P450 CLA (CYP152A2) where, despite high peroxygenase activity ( $200 \text{ min}^{-1}$ ) with myristic acid, use of  $200 \mu\text{M}$   $\text{H}_2\text{O}_2$  caused enzyme inactivation in 2-4 minutes (14). Also, using  $\text{H}_2\text{O}_2$  in a bacterial cell-based alkene-producing system may not be viable, as  $\text{H}_2\text{O}_2$  levels of only  $2 \mu\text{M}$  can cause growth inhibition of *E. coli* (15). Girhard et al. described a novel method of driving peroxygenases, using light to excite flavin cofactors with ethylenediaminetetraacetic acid (EDTA) as an electron donor. Electron transfer from reduced flavins then converts dioxygen to  $\text{H}_2\text{O}_2$ . The approach was successful with P450 CLA and P450 BS<sub>B</sub>, where near-complete conversion of  $200 \mu\text{M}$  myristic acid was observed using flavin mononucleotide (FMN) (16). This method also worked with OleT<sub>JE</sub>, where 99% of stearic acid (C18:0) substrate was oxidised, a much higher conversion rate than achieved by direct  $\text{H}_2\text{O}_2$  addition either at the start of the process, or stepwise during the reaction (13). However, this light driven approach produced low conversion levels for the shorter chain substrates lauric acid (C12:0) and myristic acid (13).

In this manuscript, we present novel alkene-producing systems in which the  $\text{H}_2\text{O}_2$ -forming alditol oxidase (AldO) from *Streptomyces coelicolor* is fused to OleT<sub>JE</sub>. AldO is a 45.1 kDa flavoprotein containing covalently bound flavin adenine dinucleotide (FAD) (17). AldO catalyses oxidation of the primary alcohol moiety of alditols using molecular oxygen, and forms the aldose product and  $\text{H}_2\text{O}_2$ . AldO oxidizes several polyol substrates, including glycerol, sorbitol and diols such as 1,2-propanediol and 1,2-hexanediol (18). Our rationale for creating the OleT<sub>JE</sub>-AldO fusion is that the  $\text{H}_2\text{O}_2$  coproduct from AldO should drive fatty acid decarboxylation by OleT<sub>JE</sub> and that the system could be regulated through measured alditol substrate additions to maintain  $\text{H}_2\text{O}_2$  concentrations compatible with efficient substrate conversion, while minimising heme destruction by excess  $\text{H}_2\text{O}_2$ . We present catalytic data showing the efficiency of the reactions, as well as using OleT<sub>JE</sub> and AldO fusion systems to identify novel products of OleT<sub>JE</sub>-dependent fatty acid oxidation.

## Materials and Methods

### Gene Cloning

The OleT<sub>JE</sub> (CYP152L1) gene from *Jeotgalicoccus* sp 8456 was codon optimised for expression in *E. coli*, synthesised (Genscript, Cherwell UK) and cloned into pET15b at NdeI/BamHI sites. The construct contains an N-terminal polyhistidine tag and a TEV cleavage site 5' to the NdeI cleavage site. The alditol oxidase (AldO) gene from *Streptomyces coelicolor* A3(2) with an N-terminal polyhistidine tag and TEV cleavage site was codon optimised for *E. coli* expression and synthesised (Genscript), prior to cloning into pET24b at NdeI/BamHI sites.

Cloning of the OleT<sub>JE</sub>-HRV3C-AldO fusion protein gene construct (linker sequence: GSGLEVLFGQPGSGGGGS with cleavage site underlined) was performed by amplification of OleT<sub>JE</sub>/AldO genes from pET15b/pET24b plasmids using primers OleT<sub>JE</sub>-For: GTATTTCCAAGGCCATATG, OleT<sub>JE</sub>-Rev: CCCCTGGAACAGAACTTCCAGACCAGAACCGGTGCGGTC CACAAC, AldO-For: GAAGTTCTGTTCCAGGGGCCGATCTGGCGGCGGC, and AldO-Rev: ATTGCGGATCCTTATCAGCCGGCCAG using Phusion High Fidelity DNA polymerase (NEB, Hitchin UK). The 5'-OleT<sub>JE</sub> and AldO-3' genes were then joined with an HRV3C cleavage site by PCR using primers

GTATTTCCAAGGCCATATG and ATTGCGGATCCTTATCAGCCGGCCAG, respectively. The OleT<sub>JE</sub>-HRV3C-AldO gene was then cloned into the pET15b-TEV plasmid at NdeI/BamHI sites (**Figure 2**).

The OleT<sub>JE</sub>- $\alpha$ helix-AldO construct was produced using a NEB HiFi DNA assembly Kit (NEB) according to manufacturer's instructions. DNA encoding the A(EAAAK)<sub>4</sub>LEA(EAAAK)<sub>4</sub>A  $\alpha$ -helix was synthesised (Eurofins, Manchester UK). The pET15b OleT<sub>JE</sub>-HRV3C-AldO plasmid was used as a template, and primers ATGTCGGATATTACCGTTACG and GGTGCGGTCCACAACTTC were used to amplify the backbone. The  $\alpha$ -helix insert was amplified with backbone overhangs of 15 bp using primers GTTGTGGACCGCACCGCTGAGGCTGCCGCTAAG and ACGTAATATCCGACATCGCTTTTGCTGCTGCTTC (**Figure 2**).

### Gene expression and protein purification

The pET15b-OleT<sub>JE</sub>, pET15b-OleT<sub>JE</sub>-HRV3C-AldO and pET15b-OleT<sub>JE</sub>- $\alpha$ helix-AldO plasmids were transformed into *E. coli* C41 (DE3) and grown in 500 ml 2xYT cultures. Cultures were shaken at 200 rpm at 37°C until an OD<sub>600</sub> of 0.5. 500  $\mu$ M  $\delta$ -aminolevulinic acid ( $\Delta$ ALA) was added and gene expression induced with 100  $\mu$ M isopropyl 1-thio- $\beta$ -D-galactopyranoside (IPTG). Temperature was decreased to 25°C for pET15b-OleT<sub>JE</sub> transformed cells, and to 20°C for cells transformed with pET15b-OleT<sub>JE</sub>-HRV3C-AldO and pET15b-OleT<sub>JE</sub>- $\alpha$ helix-AldO constructs. Cells were grown for a further 20 hours before harvesting by centrifugation at 6000 rpm at 4°C in a JLA-8.1 rotor using an Avanti J-26 XP centrifuge. Cell pellets were washed with ice-cold 100 mM KPi buffer (pH 8), centrifuged again and stored at -80°C until required for purification.

### Purification of OleT<sub>JE</sub>, OleT<sub>JE</sub>-HRV3C-AldO, OleT<sub>JE</sub>- $\alpha$ helix-AldO and P450 BM3 heme domain

OleT<sub>JE</sub> expressing cells were resuspended in 100 mM KPi, 1 M NaCl, 10% glycerol (pH 8) (buffer A). OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO expressing cells were resuspended in 100 mM KPi, 750 mM NaCl (pH 8) (buffer B). To each cell suspension, SigmaFAST Protease Inhibitor Cocktail Tablets (EDTA-Free), 100  $\mu$ g/ml DNase I and lysozyme were added (Sigma-Aldrich, Poole UK). Cell suspensions were sonicated for 40 minutes at 10 s on, 50 s off, at 40% amplitude using a Bandelin Sonopuls sonicator. The cell homogenate was then centrifuged at 20,000 rpm for 90 minutes at 4°C using a JA-25.5 rotor.

For OleT<sub>JE</sub>, 5 mM imidazole and nickel-iminodiacetic acid (Ni-IDA) chromatographic medium (Generon, Maidenhead UK) (10 ml/100 g cell pellet) was added to the supernatant and stirred overnight at 4°C. The resin was packed into a column, washed with 20 column volumes (CV) of 50 mM imidazole in buffer C (100 mM KPi, 750 mM NaCl, 10% glycerol [pH 8]) and the flow-through collected. After washing with 2 CV of 150 mM imidazole in buffer C, OleT<sub>JE</sub> protein was eluted in 175 mM imidazole in buffer C and extensively dialysed into buffer C to remove imidazole and fatty acids retained from *E. coli*. OleT<sub>JE</sub> was concentrated using a Vivaspinn (30,000 MWCO, GE Healthcare, Little Chalfont UK) and transferred into 100 mM KPi, 750 mM NaCl, 20% glycerol (pH 8) (buffer D) using a PD10 Desalting Column (GE Healthcare). The protein was then flash frozen in liquid nitrogen and stored at -80°C.

For OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO, 5 mM imidazole and Ni-IDA (5 ml/100 g) was added to cell supernatants and samples stirred at 4°C for 2 hours. The resin was washed using 20 CV of 50 mM imidazole in buffer B and the flow-through retained. Proteins were eluted with 250 mM imidazole in buffer B, and incubated overnight at 4°C with ~500 U of a S219V TEV protease variant (expressed using plasmid pRK793, Addgene plasmid no 8827) (19). The protein was exchanged into buffer B using a desalting column, and added to a bed of Ni-IDA resin. His-tag cleaved protein was eluted with 20 mM imidazole in buffer B, concentrated using a Vivaspin (30,000 MWCO) and exchanged into buffer B using a PD10 desalting column. Aliquots were frozen in liquid nitrogen and stored at -80°C.

The *Bacillus megaterium* P450 BM3 heme domain (BM3) and *Mycobacterium tuberculosis* CYP51B1 and CYP121A1 P450s were expressed and purified as described previously (20-22).

### Hydrogen peroxide tolerance of P450 enzymes

UV-visible spectroscopy was done on a Cary 60 UV-visible spectrophotometer (Agilent, Cheadle UK). Oxidative modification of OleT<sub>JE</sub> and other P450 hemes was measured at H<sub>2</sub>O<sub>2</sub> concentrations of 25, 50, 75, 125, 250, 500, 1000 and 2000  $\mu$ M in buffer C. Stock H<sub>2</sub>O<sub>2</sub> concentrations were prepared at 500x final concentration. After H<sub>2</sub>O<sub>2</sub> addition, P450 spectra were recorded every 1.5 minutes for 1 hour at 10°C. Soret peak intensity (418 nm for OleT<sub>JE</sub> and CYP51B1, 419 nm for BM3, and 417 nm for CYP121A1) was plotted against time. In each case, data were fitted accurately using a single exponential decay function.

### In vitro substrate turnover reactions with fusion proteins

Reactions were done in buffer B. To cleave OleT<sub>JE</sub>-HRV3C-AldO, HRV3C (Novagen, Feltham UK) was added at 1 U/100  $\mu$ g OleT<sub>JE</sub>-HRV3C-AldO, and incubated for 1 hour at 4°C. Proteins were exchanged into buffer B using a PD10 desalting column before starting reactions.

For reactions with OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO, 0.5 ml reactions contained 5  $\mu$ M enzyme, 500  $\mu$ M myristic acid and varying concentrations of different AldO substrates. Initial studies tested glycerol (3%, 1%, 0.1% and 0.01%), xylitol (2 mM and 10 mM) and sorbitol (2 mM and 10 mM). Reactions were incubated for 20 minutes. Time dependence studies were also done using 1% glycerol at time points of 0.5, 1, 2, 5, 10, 20 and 30 minutes. Additional reactions were done with 0.1% and 0.01% glycerol with incubations for 20 minutes. Time point experiments and reactions with 0.1% and 0.01% glycerol were done in duplicate with incubation at 27°C and shaking at 700 rpm, using 5  $\mu$ M enzyme and 500  $\mu$ M myristic acid in a 0.5 ml volume.

Reactions were stopped with 20  $\mu$ l of 37% HCl. Internal standards 1-pentadecene, palmitic acid (C16:0), 2-OH palmitic acid and 3-OH palmitic acid were added, and reactions were extracted with an equal volume of dichloromethane (DCM). The DCM layer was removed and dried with anhydrous MgSO<sub>4</sub>. DCM extract was mixed with an equal volume of N,O-bis(trimethylsilyl)trifluoroacetamide (BSTFA) containing 1% trimethylchlorosilane (TMCS) and incubated at 60°C for 45 minutes to derivatise fatty acids.

### ***In vitro* OleT<sub>JE</sub> enzymatic reactions with products**

To investigate OleT<sub>JE</sub>'s ability to further oxidize initial reaction products, 200  $\mu$ M 1-tridecene, 200  $\mu$ M  $\alpha$ -OH myristic acid or 200  $\mu$ M  $\beta$ -OH-myristic acid was incubated with either (i) 1  $\mu$ M OleT<sub>JE</sub>/500  $\mu$ M H<sub>2</sub>O<sub>2</sub> or (ii) 5  $\mu$ M OleT<sub>JE</sub>-HRV3C-AldO/1% glycerol. Reaction mixtures were incubated for 30 minutes at 27°C before acidification with 20  $\mu$ l 37% HCl, and extraction, drying and derivatisation as described previously.

### **GC/Q-TOF**

After derivatisation, 1  $\mu$ l of sample was injected onto an Agilent 7200 GC/QTOF with 7890B GC, installed with a VF-5ms 30 m x 250 mm x 0.25 mm column. The front inlet was set at 250°C, and a split ratio of 25:1 was used. Column flow was set at 1.2 ml/min, and the oven was held at 100°C for 1 minute before being ramped at 25°C/min until 200°C. Ramping was decreased to 15 ml/min until 325°C, and held for 2 minutes. Electronic ionization was used, and m/z ratios of 40-500 were recorded at 5 Hz and at 230°C. Quantitation of tridecene, myristic acid,  $\alpha$ -OH myristic acid and  $\beta$ -OH myristic acid was performed using external standard calibration curves and structural analogue internal standards.

### **Fusion protein kinetics**

To determine reaction kinetics for (i) OleT<sub>JE</sub>- $\alpha$ helix-AldO and (ii) intact and cleaved forms of OleT<sub>JE</sub>-HRV3C-AldO, decreases in myristic acid substrate concentration were plotted against reaction time. Data were fitted using a single exponential function.

## **Results**

### **Hydrogen peroxide tolerance**

The peroxygenase OleT<sub>JE</sub> uses H<sub>2</sub>O<sub>2</sub> to drive fatty acid decarboxylation. However, excess H<sub>2</sub>O<sub>2</sub> oxidatively damages heme and protein. To investigate this phenomenon, H<sub>2</sub>O<sub>2</sub> was added to OleT<sub>JE</sub> at various concentrations from 25-12000  $\mu$ M, and decreases in Soret peak intensity were monitored. To compare effects of H<sub>2</sub>O<sub>2</sub> on OleT<sub>JE</sub> with other P450s, the experiment was repeated with the fatty acid hydroxylase P450 BM3 heme domain (BM3), and the *M. tuberculosis* P450s CYP51B1 (sterol demethylase) and CYP121A1 (cyclodipeptide oxidase) (20-22). Heme modification was measured at intervals up to one hour, and at 10°C to minimise temperature-related effects on protein stability (Table 1). Figure 3 shows time-dependent changes in UV-visible spectra for OleT<sub>JE</sub> and the other P450s, demonstrating Soret band absorbance decreases indicative of heme modification. Decreases in P450 Soret peak intensity were plotted against time, and rate constants (*k*) were derived for heme oxidation processes. For OleT<sub>JE</sub>, BM3, CYP51B1 and CYP121A1, data were fitted using a single exponential function. The heme oxidation rate constant (*k*) is consistently lower for OleT<sub>JE</sub> compared to the other P450s. Differences in H<sub>2</sub>O<sub>2</sub> tolerance are most apparent at higher H<sub>2</sub>O<sub>2</sub> concentrations



(e.g.  $k = 8.51 \text{ min}^{-1}$  for OleT<sub>JE</sub> compared to  $43.25 \text{ min}^{-1}$  for BM3). Moreover, heme oxidation progresses much further for the other P450s than for OleT<sub>JE</sub> over the same time frame. These data demonstrate that OleT<sub>JE</sub> has a greater H<sub>2</sub>O<sub>2</sub> tolerance than the other bacterial P450 monooxygenases, consistent with its function as a peroxygenase. Some OleT<sub>JE</sub> heme depletion clearly occurs at 1 mM H<sub>2</sub>O<sub>2</sub>, and peroxide concentrations in this range are typically used in peroxygenase turnover experiments with fatty acids (10,23). However, more efficient fatty acid oxidation (with less heme destruction) could be achieved in a catalytic system where H<sub>2</sub>O<sub>2</sub> is produced (and enzymatically consumed) continuously at a level that is non-destructive to OleT<sub>JE</sub> heme.

### Construction of OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO fusion proteins

Two fusion proteins (OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO) were constructed by linking OleT<sub>JE</sub> to *S. coelicolor* AldO. In these systems, AldO produces H<sub>2</sub>O<sub>2</sub> from oxidation of substrates including glycerol, sorbitol and xylitol. H<sub>2</sub>O<sub>2</sub> then drives fatty acid decarboxylation by OleT<sub>JE</sub>. The OleT<sub>JE</sub>-HRV3C-AldO fusion contains a short peptide linker incorporating a HRV3C cleavage site. The protease cleavage site enables production of stoichiometric amounts of free OleT<sub>JE</sub> and AldO, allowing for comparative analysis of the activity of the separated enzymes. The OleT<sub>JE</sub>-HRV3C-AldO protein was produced at ~2.5 mg/L culture. Fusing human growth hormone (hGH) and transferrin (Tf) with a rigid helical linker was shown to be more successful for protein production by comparison to hGH/Tf fusions without a helical linker (24,25). We adopted a similar strategy to produce the OleT<sub>JE</sub>- $\alpha$ helix-AldO fusion enzyme, which was also purified at ~2.5 mg/L. **Figure 4** shows SDS-PAGE gels of both constructs following purification by Ni-IDA chromatography, TEV cleavage of the polyhistidine tag, and separation of the untagged fusion protein by reverse Ni-IDA chromatography. **Figure 4A** also shows OleT<sub>JE</sub>-HRV3C-AldO following inter-domain cleavage using HRV3C protease.

### Driving fatty acid decarboxylation with different AldO substrates

Reactions were set up with 5  $\mu\text{M}$  OleT<sub>JE</sub>-HRV3C-AldO, 0.5 mM myristic acid and different AldO substrates (glycerol, sorbitol and xylitol). An additional reaction was set up with 500  $\mu\text{M}$  H<sub>2</sub>O<sub>2</sub> and 5  $\mu\text{M}$  OleT<sub>JE</sub>-HRV3C-AldO. A control reaction without substrate was also set up, including 40 units of catalase to remove any H<sub>2</sub>O<sub>2</sub>. **Table 2** shows that all AldO substrates gave high levels of turnover, facilitating myristic acid oxidation levels of 91-95%, compared to 87% in the case where H<sub>2</sub>O<sub>2</sub> was added directly to substrate-bound OleT<sub>JE</sub>. Glycerol gave marginally the highest levels of substrate oxidation, and was used as the AldO substrate in further experiments.

### Time-dependent oxidation of fatty acids using fusion proteins

Reactions were set up with 1% glycerol, 0.5 mM myristic acid, and 5  $\mu\text{M}$  of (i) intact OleT<sub>JE</sub>-HRV3C-AldO, (ii) cleaved OleT<sub>JE</sub>-HRV3C-AldO or (iii) OleT<sub>JE</sub>- $\alpha$ helix-AldO. Myristic acid concentration was plotted against time, and rate constants ( $k$ ) determined for fatty acid conversion. The highest rate was obtained for OleT<sub>JE</sub>- $\alpha$ helix-AldO ( $0.48 \pm 0.02 \text{ min}^{-1}$ ). OleT<sub>JE</sub>-HRV3C-AldO and the HRV3C-cleaved



forms showed conversion rates of  $0.38 \pm 0.05 \text{ min}^{-1}$  and  $0.15 \pm 0.01 \text{ min}^{-1}$ , respectively, indicating that the fused enzymes have higher activity than their separate entities at the same concentration. **Figure 5** shows GC-MS analysis of myristic acid products formed by the OleT<sub>JE</sub>- $\alpha$ helix-AldO fusion enzyme.

Product formation over time was also monitored for the conditions (i)-(iii). In all cases, tridecene was the major product after 20 minutes (**Table 3**). However, for both intact and cleaved OleT<sub>JE</sub>-HRV3C-AldO, 3-OH myristic acid is initially produced at high levels, and is the major product at 1 minute. Levels of 2-OH and 3-OH myristic acid decreased after 2 minutes in all three conditions. We hypothesised that, with continued production of H<sub>2</sub>O<sub>2</sub> by AldO-dependent glycerol oxidation, the hydroxylated fatty acids are further converted into other products. This finding indicates that the OleT<sub>JE</sub> protein constructs used remain active throughout the assays and can bind and further oxidise the primary reaction products.

### Optimising glycerol concentration for OleT<sub>JE</sub> activity

To establish suitable glycerol concentrations for OleT<sub>JE</sub>-AldO fusion proteins, reactions were also performed with 0.1 and 0.01% glycerol, and stopped after 20 minutes. **Table 3** compares the percentage conversion of myristic acid for (i) intact OleT<sub>JE</sub>-HRV3C-AldO, (ii) cleaved OleT<sub>JE</sub>-HRV3C-AldO, and (iii) OleT<sub>JE</sub>- $\alpha$ helix-AldO, as well as the concentrations of tridecene, 2-OH myristic acid and 3-OH myristic acid products formed. For all proteins, there was little activity at 0.01% glycerol. However, hydroxylated products were formed at higher levels than alkene at low glycerol concentrations. At 0.1% glycerol, levels of 2-OH and 3-OH myristic acid are higher than at 1% glycerol. This may occur due to limited enzymatic activity at the lower glycerol concentration, resulting in less efficient secondary oxidation of the primary hydroxylated reaction products.

For intact OleT<sub>JE</sub>-HRV3C-AldO, the 1% glycerol condition yielded a 15% increase in total turnover and 9% increase in alkene production compared to 0.1% glycerol. There was little difference in total turnover and alkene levels for the intact and HRV3C-cleaved forms of OleT<sub>JE</sub>-HRV3C-AldO, suggesting that proteolytic cleavage is effective and does not inactivate enzymatic functions.

### Secondary turnover of 2-OH myristic acid and 3-OH myristic acid products

Time-course reactions with OleT<sub>JE</sub>-AldO fusion proteins showed that levels of 2-OH and 3-OH myristic acid decreased over time (from ~2 minutes onwards). This suggested that OleT<sub>JE</sub> can convert these hydroxylated fatty acids to further products (**Figure 6**). 0.5 ml reactions were set up with 5  $\mu\text{M}$  OleT<sub>JE</sub>-HRV3C-AldO, 1% glycerol and 200  $\mu\text{M}$  of either 2-OH or 3-OH myristic acid, or using 1  $\mu\text{M}$  OleT<sub>JE</sub>/500  $\mu\text{M}$  H<sub>2</sub>O<sub>2</sub>. Previous studies on the *S. coelicolor* AldO enzyme reported a  $k_{\text{cat}} = 17 \text{ s}^{-1}$  and a  $k_{\text{cat}}/K_{\text{m}} = 1.2 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$  for sorbitol, and values of  $k_{\text{cat}} = 13 \text{ s}^{-1}$  and  $k_{\text{cat}}/K_{\text{m}} = 4.1 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$  for xylitol at pH 7.5 (26); and a  $k_{\text{cat}}/K_{\text{m}} = 2.1 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$  for glycerol at pH 9.0 (27). Given the similar extents of oxidation of these different substrates by the OleT<sub>JE</sub>-HRV3C-AldO fusion enzyme (**Table 2**), it is likely that the kinetics of this fusion enzyme are similar to those reported previously.

Reactions were incubated for 30 minutes with mixing at 700 rpm at 27°C. GC/Q-TOF chromatograms were compared to those of control samples without enzyme. Data showed that both 2-OH and 3-OH myristic acid were used effectively as OleT<sub>JE</sub> substrates, with 3-OH myristic acid reactions showing almost complete turnover (**Figure 7**). For the 2-OH myristic acid reaction, two products were identified. One of these products is tridec-1-en-1-ol, indicating that OleT<sub>JE</sub> can decarboxylate 2-OH myristic acid. The ion spectrum of the second product indicates that it is 2-hydroxytetradec-2-enoic acid. This demonstrates that OleT<sub>JE</sub> can desaturate 2-OH myristic acid at the C<sub>α</sub>-C<sub>β</sub> bond. For reactions with 3-OH myristic acid, the product is 3,4 dihydroxytetradec-2-enoic acid, indicating that OleT<sub>JE</sub> can further hydroxylate as well as desaturate 3-OH myristic acid.

## Discussion

The *Jeotgalicoccus* sp. OleT<sub>JE</sub> is an efficient P450 peroxygenase that uses H<sub>2</sub>O<sub>2</sub> to oxidatively decarboxylate fatty acids to form terminal alkenes. This peroxide shunt mechanism (**Figure 1**) to form compound 0 is inefficient in many P450s, including BM3, causing heme oxidative damage and giving low product formation. Efforts have been made to enhance peroxide shunt efficiency through protein engineering of BM3 and the camphor hydroxylase P450cam, although improvements were not substantial (28,29). Our data (using heme stability analysis) show that OleT<sub>JE</sub> is relatively tolerant to H<sub>2</sub>O<sub>2</sub>, and clearly more resistant to H<sub>2</sub>O<sub>2</sub>-mediated heme oxidation than are the BM3, CYP121A1 and CYP51B1 P450s (**Figure 3, Table 1**). Even after incubation of OleT<sub>JE</sub> with 1 mM H<sub>2</sub>O<sub>2</sub>, its Fe<sup>II</sup>-CO spectrum remains almost completely in the P450 (A<sub>max</sub> = 449 nm) form, indicative of negligible loss of its cysteine thiolate heme ligand. However, despite its evolution into a peroxygenase, it remains somewhat susceptible to heme modification at higher concentrations of H<sub>2</sub>O<sub>2</sub> (e.g. 1-2 mM), although the enzyme should not encounter such elevated levels of H<sub>2</sub>O<sub>2</sub> in the bacterial cell.

In this study, we report the production and catalytic properties of AldO-OleT<sub>JE</sub> fusion systems. AldO produces H<sub>2</sub>O<sub>2</sub> as a co-product from oxidation of substrates including glycerol, sorbitol and xylitol. The fused OleT<sub>JE</sub> uses the H<sub>2</sub>O<sub>2</sub> to drive decarboxylation and hydroxylation of myristic acid, a good substrate for OleT<sub>JE</sub> (7). In a system where OleT<sub>JE</sub> and AldO proteins are fused by a linker containing a HRV3C cleavage site (OleT<sub>JE</sub>-HRV3C-AldO), addition of 1% glycerol gave an 8% increase in myristic acid conversion compared to addition of 0.5 mM H<sub>2</sub>O<sub>2</sub> (following a 20 minute incubation). Glycerol is abundant and inexpensive (30), and is formed as a by-product in the production of biofuels, with associated disposal costs (31). Glycerol would thus be an ideal substrate for cheap and efficient alkene production using AldO-OleT<sub>JE</sub> fusion enzymes.

The kinetics of myristic acid oxidation and product formation were measured for (i) OleT<sub>JE</sub>-HRV3C-AldO, (ii) OleT<sub>JE</sub>-αhelix-AldO, a construct in which OleT<sub>JE</sub> and AldO are fused by a non-cleavable alpha helical linker, and (iii) the separated OleT<sub>JE</sub> and AldO entities produced stoichiometrically by HRV3C-dependent cleavage of the OleT<sub>JE</sub>-HRV3C-AldO linker. The fused proteins showed higher rates of myristic acid turnover than their separated entities, with 97% myristic acid turnover catalysed by OleT<sub>JE</sub>-αhelix-AldO after 20 minutes. Fusion of partner domains thus enhances product formation in comparison to equal amounts of isolated OleT<sub>JE</sub> and AldO. The ability to co-express functional and stable OleT<sub>JE</sub>/AldO fusion enzymes is also advantageous in producing catalytically self-sufficient enzymes. The proximity of these domains in the fusion enzymes may enhance activity through more

efficient channelling of H<sub>2</sub>O<sub>2</sub> between the enzyme active sites, and in a cellular context this could be important in ensuring that fatty acid decarboxylation/hydroxylation is well coupled to H<sub>2</sub>O<sub>2</sub> production. The regulated and continuous glycerol-dependent production of H<sub>2</sub>O<sub>2</sub> from the OleT<sub>JE</sub>/AldO fusion enzymes described here is likely conducive to more efficient fatty acid oxidation. Previous studies of bacterial peroxygenases have also used such continuous H<sub>2</sub>O<sub>2</sub> production strategies (e.g. a light-driven H<sub>2</sub>O<sub>2</sub>-producing system) (13,32). Our ongoing studies involve analysis of product formation from the OleT<sub>JE</sub>-AldO fusion enzymes expressed in *E. coli* cells to establish reaction efficiency and products formed.

Importantly, time-dependent studies of myristic acid product formation from OleT<sub>JE</sub>-AldO fusion enzymes indicated the early formation and then decrease in concentration of 2-OH and 3-OH myristic acid products. Reactions done at lower glycerol concentration (0.1%) produced higher levels of hydroxylated myristic acid, possibly due to decreased levels of secondary turnover of the hydroxylated products through weaker enzyme activity at lower glycerol concentrations. Incubation of 2-OH myristic acid with OleT<sub>JE</sub>/H<sub>2</sub>O<sub>2</sub>, or with OleT<sub>JE</sub>-HRV3C-AldO and 1% glycerol, led to further turnover of 2-OH myristic acid and formation of 2-hydroxytetradec-2-enoic acid and tridec-1-en-1-ol, indicating that OleT<sub>JE</sub> can decarboxylate 2-OH myristic acid as well as catalysing desaturation of this substrate. Desaturation reactions are among the wide repertoire of molecular transformations catalysed by P450s. These may occur either due to (i) imperfect positioning of the substrate carbon radical that forms following hydrogen abstraction to form P450 compound II (Fe<sup>IV</sup>-OH), leading to an electron transfer from the carbon radical to the iron, forming a carbocation. Loss of the proton adjacent to the carbocation then results in double bond formation; or (ii) compound II not recombining with the substrate carbon radical, and instead abstracting a hydrogen atom from a carbon adjacent to the radical to form the double bond (33). Parallel reactions also revealed extensive conversion of 3-OH myristic acid, and formation of 3,4 dihydroxytetradec-2-enoic acid. These data demonstrate that OleT<sub>JE</sub> can di-hydroxylate the fatty acid, in addition to desaturating the C<sub>α</sub>-C<sub>β</sub> bond. OleT<sub>JE</sub> was previously shown to produce 2-alkanones from reactions with palmitic acid and stearic acid (C16:0 and 18:0) (34). However, we provide the first examples for the secondary turnover of primary OleT<sub>JE</sub> reaction products with myristic acid.

In conclusion, we describe the development of novel alditol oxidase-peroxygenase (AldO-OleT<sub>JE</sub>) fusion enzymes that enable production of alkenes using glycerol and other alditol substrates. Further, we use these fusion systems to characterize previously undescribed decarboxylated, hydroxylated and desaturated products formed from primary reaction products (2- and 3-hydroxy myristic acid). These data provide new insights into the catalytic repertoire of OleT<sub>JE</sub>, as well as providing efficient new fusion enzyme systems for alkene production.

## Author Contributions statement

AWM, RTB and DL conceived and supervised the study. AWM, SM, K-LT, RTB and KJM designed experiments. SM, K-LT, KJM and NJR performed experiments. SM, KJM, DL and K-LT analysed data. SM, AWM and KJM wrote the manuscript. All authors made manuscript revisions.

## REFERENCES

1. Luthra, A., Denisov, I. G., and Sligar, S. G. (2011) Spectroscopic features of cytochrome P450 reaction intermediates. *Arch Biochem Biophys* **507**, 26-35
2. Sligar, S. G., and Gunsalus, I. C. (1979) Proton coupling in the cytochrome P-450 spin and redox equilibria. *Biochemistry* **18**, 2290-2295
3. Munro, A. W., Girvan, H. M., and McLean, K. J. (2007) Variations on a (t)heme - novel mechanisms, redox partners and catalytic functions. *Nat Prod Rep* **24**, 585-609
4. Denisov, I. G., Makris, T. M., Sligar, S. G., and Schlichting, I. (2005) Structure and chemistry of cytochrome P450. *Chem Rev* **105**, 2253-2277
5. Rittle, J., and Green, M. T. (2010) Cytochrome P450 compound I: capture, characterization, and C-H bond activation kinetics. *Science* **330**, 933-937
6. Rude, M. A., Baron, T. S., Brubaker, S., Alibhai, M., Del Cardayre, S. B., and Schirmer, A. (2011) Terminal olefin (1-alkene) biosynthesis by a novel P450 fatty acid decarboxylase. *Appl Environ Microbiol* **77**, 1718-1727
7. Liu, Y., Wang, C., Yan, J., Zhang, W., Guan, W., Lu, X., and Li, S. (2014) Hydrogen peroxide-independent production of alpha-alkenes by OleT<sub>JE</sub> P450 fatty acid decarboxylase. *Biotechnol Biofuels* **7**, 28
8. Wang, W., and Lu, X. (2013) Microbial Synthesis of Alka(e)nes. *Front Bioeng Biotechnol* **1**, 10
9. Herman, N. A., and Zhang, W. (2016) Enzymes for fatty acid-based hydrocarbon biosynthesis. *Curr Opin Chem Biol* **35**, 22-28
10. Belcher, J., McLean, K. J., Matthews, S., Woodward, L. S., Fisher, K., Rigby, S. E., Nelson, D. R., Potts, D., Baynham, M. T., Parker, D. A., Leys, D., and Munro, A. W. (2014) Structure and biochemical properties of the alkene producing cytochrome P450 OleT<sub>JE</sub> (CYP152L1) from the *Jeotgalicoccus* sp. 8456 bacterium. *J Biol Chem* **289**, 6535-6550
11. Grant, J. L., Hsieh, C. H., and Makris, T. M. (2015) Decarboxylation of fatty acids to terminal alkenes by cytochrome P450 compound I. *J Am Chem Soc* **137**, 4940-4943
12. Grant, J. L., Mitchell, M. E., and Makris, T. M. (2016) Catalytic strategy for carbon-carbon bond scission by the cytochrome P450 OleT. *Proc Natl Acad Sci U S A* **113**, 10049-10054
13. Zachos, I., Gassmeyer, S. K., Bauer, D., Sieber, V., Hollmann, F., and Kourist, R. (2015) Photobiocatalytic decarboxylation for olefin synthesis. *Chem Commun (Camb)* **51**, 1918-1921
14. Girhard, M., Schuster, S., Dietrich, M., Durre, P., and Urlacher, V. B. (2007) Cytochrome P450 monooxygenase from *Clostridium acetobutylicum*: a new alpha-fatty acid hydroxylase. *Biochem Biophys Res Commun* **362**, 114-119
15. Seaver, L. C., and Imlay, J. A. (2001) Hydrogen peroxide fluxes and compartmentalization inside growing *Escherichia coli*. *J Bacteriol* **183**, 7182-7189
16. Girhard, M., Kunigk, E., Tihovsky, S., Shumyantseva, V. V., and Urlacher, V. B. (2013) Light-driven biocatalysis with cytochrome P450 peroxxygenases. *Biotechnol Appl Biochem* **60**, 111-118
17. Heuts, D. P., van Hellemond, E. W., Janssen, D. B., and Fraaije, M. W. (2007) Discovery, characterization, and kinetic analysis of an alditol oxidase from *Streptomyces coelicolor*. *J Biol Chem* **282**, 20283-20291
18. van Hellemond, E. W., Vermote, L., Koolen, W., Sonke, T., Zandvoort, E., Heuts, D. P. H. M., Janssen, D. B., and Fraaije, M. W. (2009) Exploring the biocatalytic scope of alditol oxidase from *Streptomyces coelicolor*. *Adv Synth Catal* **351**, 1523-1530

19. Kapust, R. B., Tozser, J., Fox, J. D., Anderson, D. E., Cherry, S., Copeland, T. D., and Waugh, D. S. (2001) Tobacco etch virus protease: mechanism of autolysis and rational design of stable mutants with wild-type catalytic proficiency. *Protein Eng* **14**, 993-1000
20. Butler, C. F., Peet, C., Mason, A. E., Voice, M. W., Leys, D., and Munro, A. W. (2013) Key mutations alter the cytochrome P450 BM3 conformational landscape and remove inherent substrate bias. *J Biol Chem* **288**, 25387-25399
21. McLean, K. J., Carroll, P., Lewis, D. G., Dunford, A. J., Seward, H. E., Neeli, R., Cheesman, M. R., Marsollier, L., Douglas, P., Smith, W. E., Rosenkrands, I., Cole, S. T., Leys, D., Parish, T., and Munro, A. W. (2008) Characterization of active site structure in CYP121. A cytochrome P450 essential for viability of *Mycobacterium tuberculosis*. *J Biol Chem* **283**, 33406-33416
22. McLean, K. J., Warman, A. J., Seward, H. E., Marshall, K. R., Girvan, H. M., Cheesman, M. R., Waterman, M. R., and Munro, A. W. (2006) Biophysical characterization of the sterol demethylase P450 from *Mycobacterium tuberculosis*, its cognate ferredoxin, and their interactions. *Biochemistry* **45**, 8427-8443
23. Hsieh, C. H., and Makris, T. M. (2016) Expanding the substrate scope and reactivity of cytochrome P450 OleT. *Biochem. Biophys. Res. Commun.* **476**, 462-466
24. Amet, N., Lee, H.-F., and Shen, W.-C. (2009) Insertion of the designed helical linker led to increased expression of Tf-based fusion proteins. *Pharm Res* **26**, 523-528
25. Chen, X., Zaro, J., and Shen, W.-C. (2013) Fusion protein linkers: Property, design and functionality. *Adv Drug Deliv Rev* **65**, 1357-1369
26. Heuts, D. P. H. M., van Hellemond, E. W., Janssen, D. B., and Fraaije, M. W. (2007) Discovery, characterization, and kinetic analysis of an alditol oxidase from *Streptomyces coelicolor*. *J Biol Chem* **282**, 20283-20281
27. Gerstenbruch, S., Wulf, H., Mussmann, N., O'Connell, T., Maurer, K.-H., and Bornscheuer, U. T. (2012) Asymmetric synthesis of D-glyceric acid by an alditol oxidase and directed evolution for enhanced oxidative activity towards glycerol. *Appl. Microbiol. Biotechnol.* **96**, 1243-1252
28. Joo, H., Lin, Z., and Arnold, F. H. (1999) Laboratory evolution of peroxide-mediated cytochrome P450 hydroxylation. *Nature* **399**, 670-673
29. Li, Q. S., Ogawa, J., and Shimizu, S. (2001) Critical role of the residue size at position 87 in H<sub>2</sub>O<sub>2</sub>- dependent substrate hydroxylation activity and H<sub>2</sub>O<sub>2</sub> inactivation of cytochrome P450BM-3. *Biochem Biophys Res Commun* **280**, 1258-1261
30. Mattam, A. J., Clomburg, J. M., Gonzalez, R., and Yazdani, S. S. (2013) Fermentation of glycerol and production of valuable chemical and biofuel molecules. *Biotechnol Lett* **35**, 831-842
31. Yazdani, S. S., and Gonzalez, R. (2007) Anaerobic fermentation of glycerol: a path to economic viability for the biofuels industry. *Curr Opin Biotechnol* **18**, 213-219
32. Amaya, J. A., Rutland, C. D., and Makris, T. M. (2016) Mixed regiospecificity compromises alkene synthesis by a cytochrome P450 peroxygenase from *Methylobacterium populi*. *J Inorg Biochem* **158**, 11-16.
33. Ortiz de Montellano, P. R. (2015) Substrate oxidation by cytochrome P450 enzymes. Pp. 111-176 in *Cytochrome P450: Structure, Mechanism and Biochemistry*, 4<sup>th</sup> Ed. Ed. Ortiz de Montellano, P.R. Published by Springer International Publishing Switzerland.



34. Dennig, A., Kuhn, M., Tassoti, S., Thiessenhusen, A., Gilch, S., Bulter, T., Haas, T., Hall, M., and Faber, K. (2015) Oxidative decarboxylation of short-chain fatty acids to 1-alkenes. *Angew Chem Int Ed Engl* **54**, 8819-8822
35. Groves, J. T. (2006) High-valent iron in chemical and biological oxidations. *J Inorg Biochem* **100**, 434-447
36. Fisher, M. T., and Sligar, S. G. (1985) Control of heme protein redox potential and reduction rate: linear free energy relation between potential and ferric spin state equilibrium. *J Am Chem Soc* **107**, 5018-5019

## FIGURES

**Figure 1. P450 catalytic cycle and active site structure of OleT<sub>JE</sub>.** Panel A shows a cytochrome P450 catalytic cycle, illustrating how OleT<sub>JE</sub> utilises the peroxide shunt pathway to catalyse fatty acid decarboxylation to form terminal alkenes. In its ferric resting state, the OleT<sub>JE</sub> heme iron is axially co-ordinated by cysteine thiolate and a water molecule. Upon addition of fatty acid (a), the axial water molecule is displaced, and the heme iron converts from a low spin (LS) to a high spin (HS) state (2). Addition of H<sub>2</sub>O<sub>2</sub> to OleT<sub>JE</sub> initiates the peroxide shunt, forming a ferric-hydroperoxo intermediate (Compound 0, reaction indicated by the arrow crossing the cycle). Protonation and dehydration of Compound 0 (f), yields a reactive ferryl (Fe<sup>IV</sup>)-oxo, porphyrin radical cation (Compound I), which has been observed in OleT<sub>JE</sub> using single turnover stopped-flow absorption spectroscopy (11). OleT<sub>JE</sub> then catalyses either decarboxylation of the fatty acid (g-i) to form terminal alkene (the major reaction), or hydroxylates the substrate at the  $\alpha$ - or  $\beta$ -position (not shown). Hydroxylation involves Compound I-mediated hydrogen abstraction from a substrate CH group (forming Compound II, Fe<sup>IV</sup>-OH), and hydroxylation by radical rebound with restoration of the ferric heme (35). However, decarboxylation likely occurs through the further abstraction of an electron from the substrate radical by Compound II (h), producing an unstable substrate carbocation or diradical that decarboxylates to generate the alkene (12). The alkene product then dissociates, releasing CO<sub>2</sub> (i), and a protonation step restores the ferric heme iron to its resting, water ligated form. OleT<sub>JE</sub> can also function using redox partner systems, in which case two electron transfer steps and one protonation step are required to convert the substrate-bound, ferric species to compound 0 [(b)-(e)] (34,36). Steps (f)-(i) then occur as described above following the peroxide shunt process (3). Panel B shows the active site of OleT<sub>JE</sub> bound to arachidic acid (C20:0), with substrate shown in orange (10) (PDB 4L40). The heme is coloured in blue, and is proximally coordinated to Cys365 thiolate. Also shown is arachidic acid (orange) and the interaction between its carboxylate and Arg245.

**Figure 2. Schematic of OleT<sub>JE</sub>-AldO constructs.** Panel A shows how an OleT<sub>JE</sub>-AldO fusion system can use *S. coelicolor* AldO to oxidise glycerol (or another alditol substrate), forming H<sub>2</sub>O<sub>2</sub> as a co-product. H<sub>2</sub>O<sub>2</sub> is then used by OleT<sub>JE</sub> to decarboxylate fatty acids such as myristic acid, producing terminal alkenes as the major product. Terminal alkenes have important potential industrial uses, including as “drop-in” biofuels. Panel B shows the fusion constructs used in this work. (i) OleT<sub>JE</sub>-HRV3C-AldO. OleT<sub>JE</sub> and AldO are fused by a HRV3C-cleavable linker, enabling stoichiometric production of OleT<sub>JE</sub> and AldO, as shown in (ii). (iii) OleT<sub>JE</sub>- $\alpha$ helix-AldO. In this construct OleT<sub>JE</sub> and AldO are linked by an A(EAAK)<sub>4</sub>LEA(EAAK)<sub>4</sub>A  $\alpha$ -helix.



**Figure 3. P450 H<sub>2</sub>O<sub>2</sub> tolerance.** Panels A-D show the UV-visible spectra of OleT<sub>JE</sub> (A), BM3 heme domain (B), CYP51B1 (C), and CYP121A1 (D), before (thick solid line) and after the addition of 1 mM H<sub>2</sub>O<sub>2</sub>. Proteins are all at a concentration of 5  $\mu$ M. These spectra show heme absorbance decrease due to H<sub>2</sub>O<sub>2</sub>-mediated oxidation of the prosthetic group, sampled over 1 hour at 1.5 minute intervals. Rate constants for heme oxidation were obtained for the four proteins at H<sub>2</sub>O<sub>2</sub> concentrations between 0.025-2 mM H<sub>2</sub>O<sub>2</sub> (see Table 1). At 1 mM H<sub>2</sub>O<sub>2</sub>, the greater stability of OleT<sub>JE</sub> enzyme compared to the other (non-peroxygenase) P450s is clear, with much greater heme oxidation and absorbance loss observed for BM3 heme domain, CYP51B1 and CYP121A1 compared to OleT<sub>JE</sub> under the same conditions.

**Figure 4: SDS-PAGE analysis of purified OleT<sub>JE</sub> fusion proteins.** Panel A shows intact OleT<sub>JE</sub>-HRV3C-AldO (i) and cleaved OleT<sub>JE</sub>-HRV3C-AldO (ii), following incubation of the fusion protein with HRV3C protease for 1 hour at 4°C. Panel B shows purified OleT<sub>JE</sub>- $\alpha$ helix-AldO (iii). Samples were resolved by SDS-PAGE using a 10% polyacrylamide gel. The first lane in each case shows protein markers (NEB Broad Range Protein Ladder).

**Figure 5: GC/MS chromatogram showing products formed by the OleT<sub>JE</sub>- $\alpha$ helix-AldO fusion protein with myristic acid substrate.** The reaction shown was performed at 27°C with shaking at 700 rpm, and with 0.5 mM myristic acid, 5  $\mu$ M OleT<sub>JE</sub>- $\alpha$ helix-AldO fusion protein and 0.1% glycerol. The black chromatogram shows reaction products formed with the reaction stopped at 20 minutes. The red chromatogram shows a control reaction containing the same components, but also including catalase (40 units) to remove H<sub>2</sub>O<sub>2</sub>. The peaks relate to the following compounds: i) tridecene, ii) pentadecene (internal standard), iii) myristic acid, iv) 2-OH myristic acid, v) 3-OH myristic acid, and vi) palmitic acid (internal standard).

**Figure 6. Time dependence of myristic acid conversion and product formation at 1% glycerol.** Panels A, B and C show the extents of myristic acid conversion using GC/MS data collected following stopping the reaction immediately after its initiation (~10 s), and then after 0.5, 1, 2, 5, 10 and 20 minutes for intact OleT<sub>JE</sub>-HRV3C-AldO, cleaved OleT<sub>JE</sub>-HRV3C-AldO and OleT<sub>JE</sub>- $\alpha$ helix-AldO, respectively. Panels D, E and F show concentrations of tridecene, 2-OH myristic acid and 3-OH myristic acid in samples taken over 20 minutes for the same enzymes. Duplicate samples were tested and standard errors are shown.

**Figure 7. Secondary turnover of primary hydroxylated products from OleT<sub>JE</sub>-HRV3C-AldO fusion protein-mediated myristic acid oxidation.** Panel A shows chromatograms from a reaction (black) with 5  $\mu$ M OleT<sub>JE</sub>-HRV3C-AldO fusion protein, 200  $\mu$ M 2-OH myristic acid and 500  $\mu$ M H<sub>2</sub>O<sub>2</sub>, as well as a control sample with no enzyme (red). The reaction shows a decrease in 2-OH myristic acid (ii) and the appearance of peaks (i) and (iii). The ion spectra identify these molecules as (i) tridec-1-en-1-ol and (iii) 2-hydroxytetradec-2-enoic acid. Panel B shows reaction (black) and control (red) with 200  $\mu$ M 3-OH myristic acid (same conditions as for panel A). The data show the disappearance of 3-OH myristic acid (iv), and the appearance of peak (v), which is identified as 3,4 dihydroxytetradec-2-enoic acid.

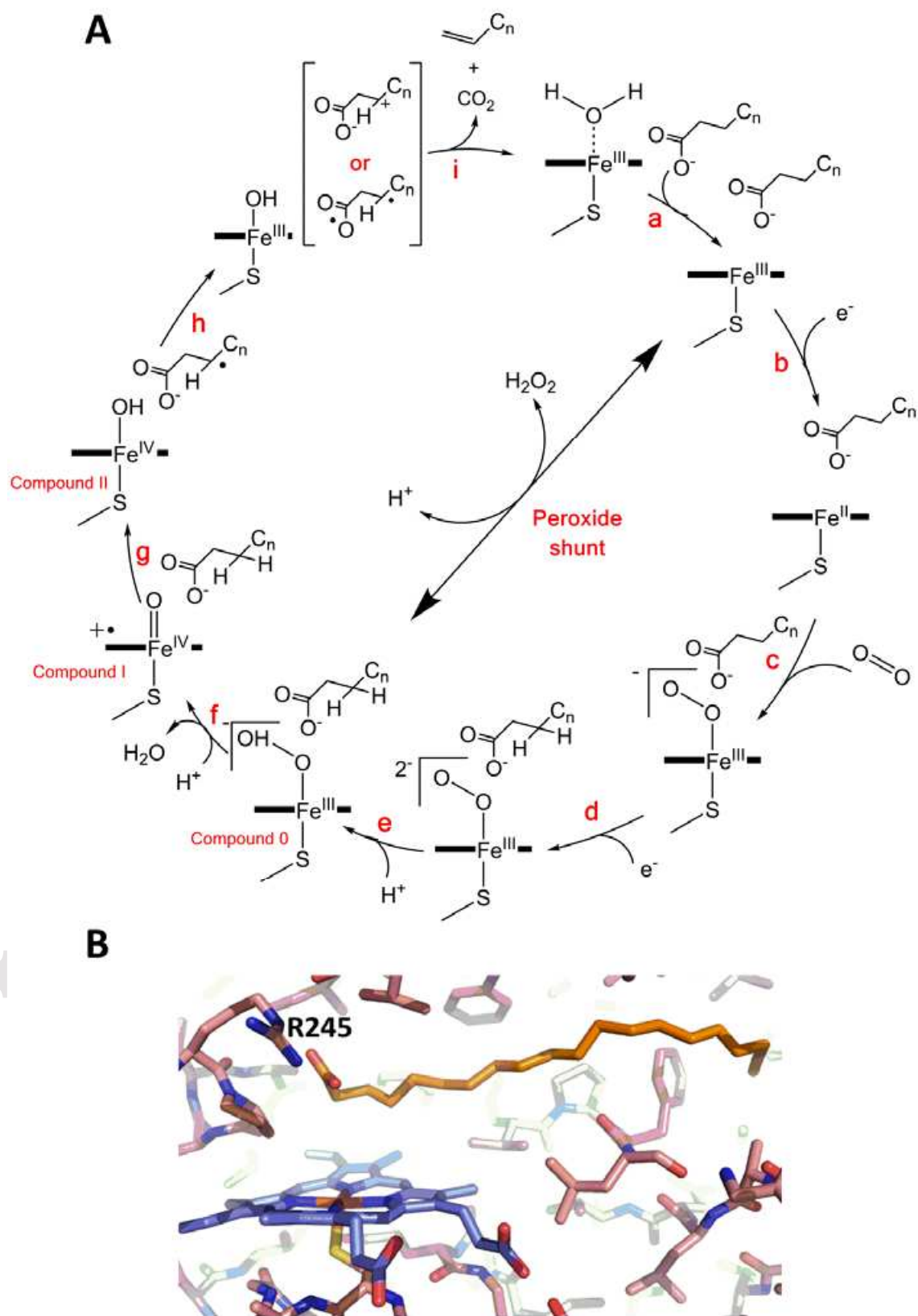


Figure 1

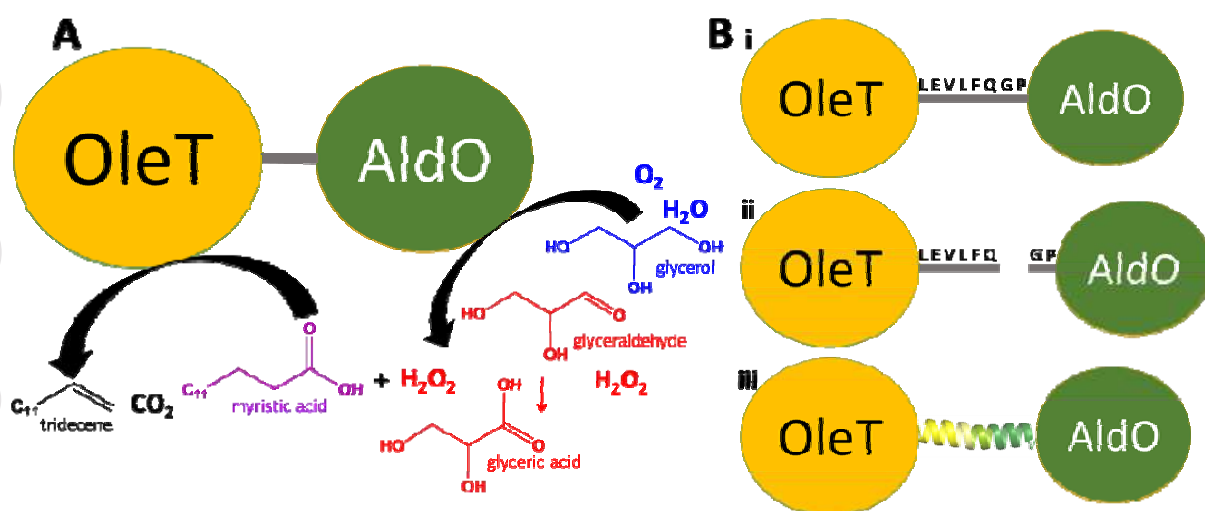


Figure 2

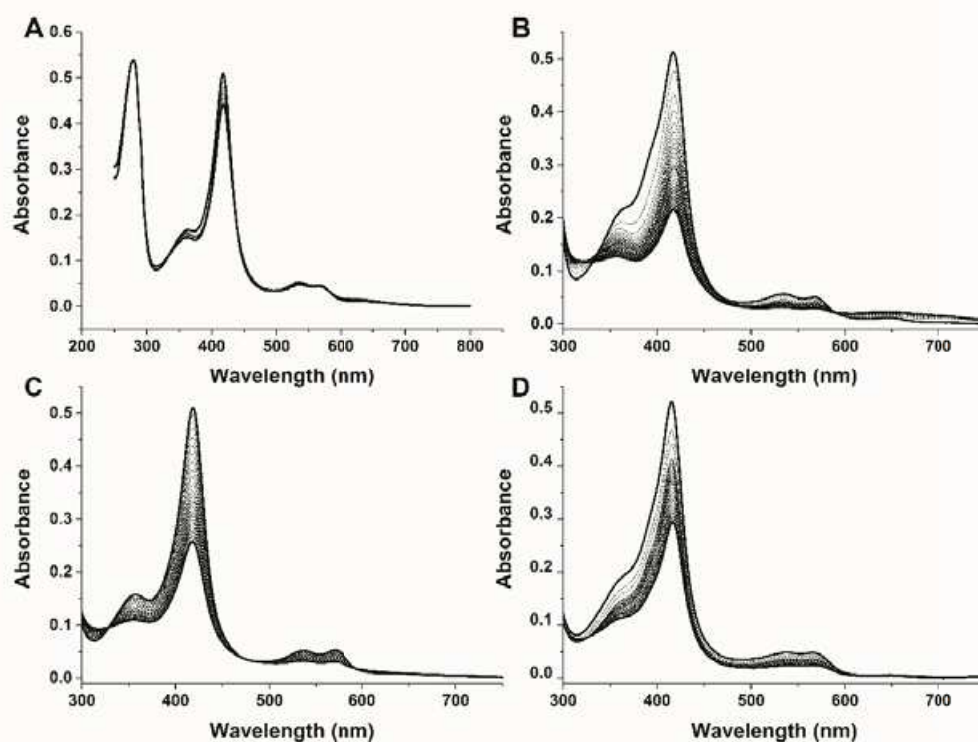


Figure 3

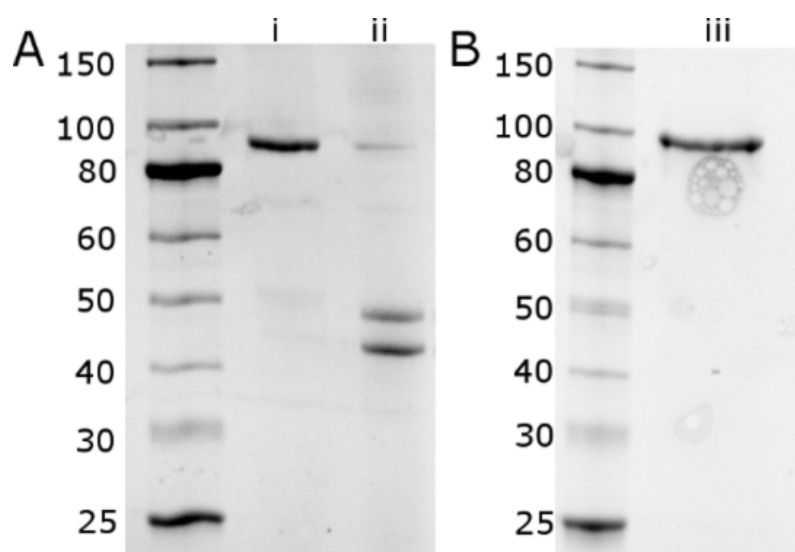


Figure 4

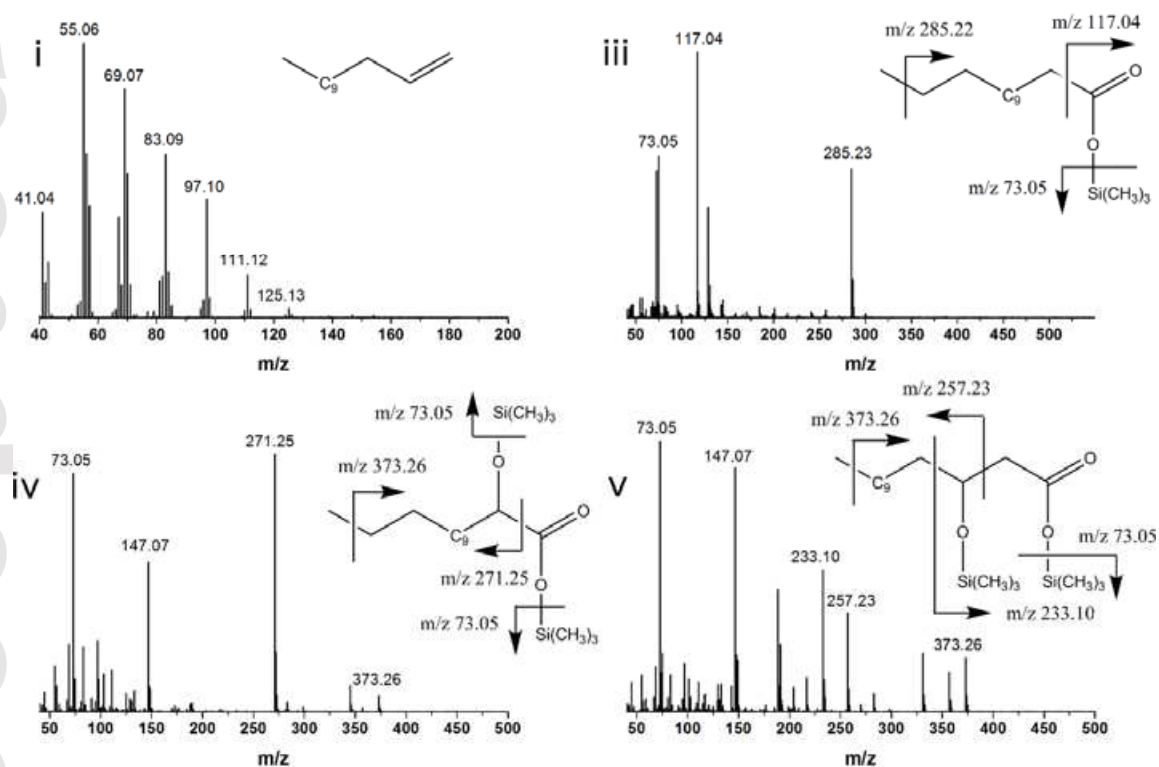
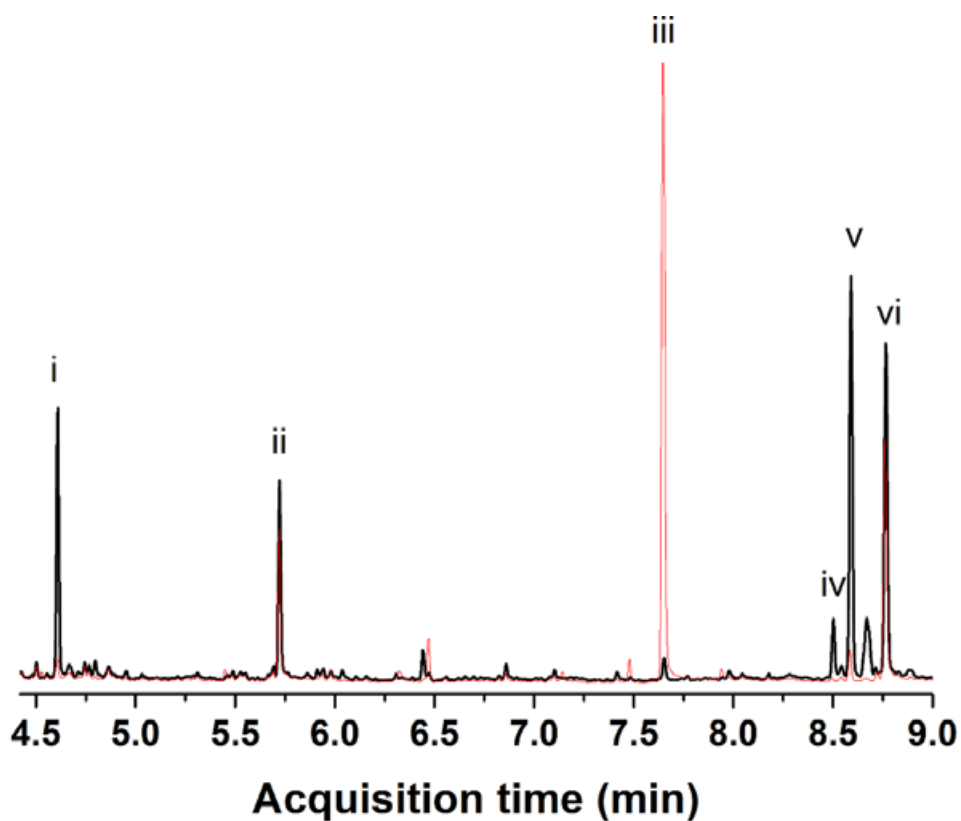


Figure 5

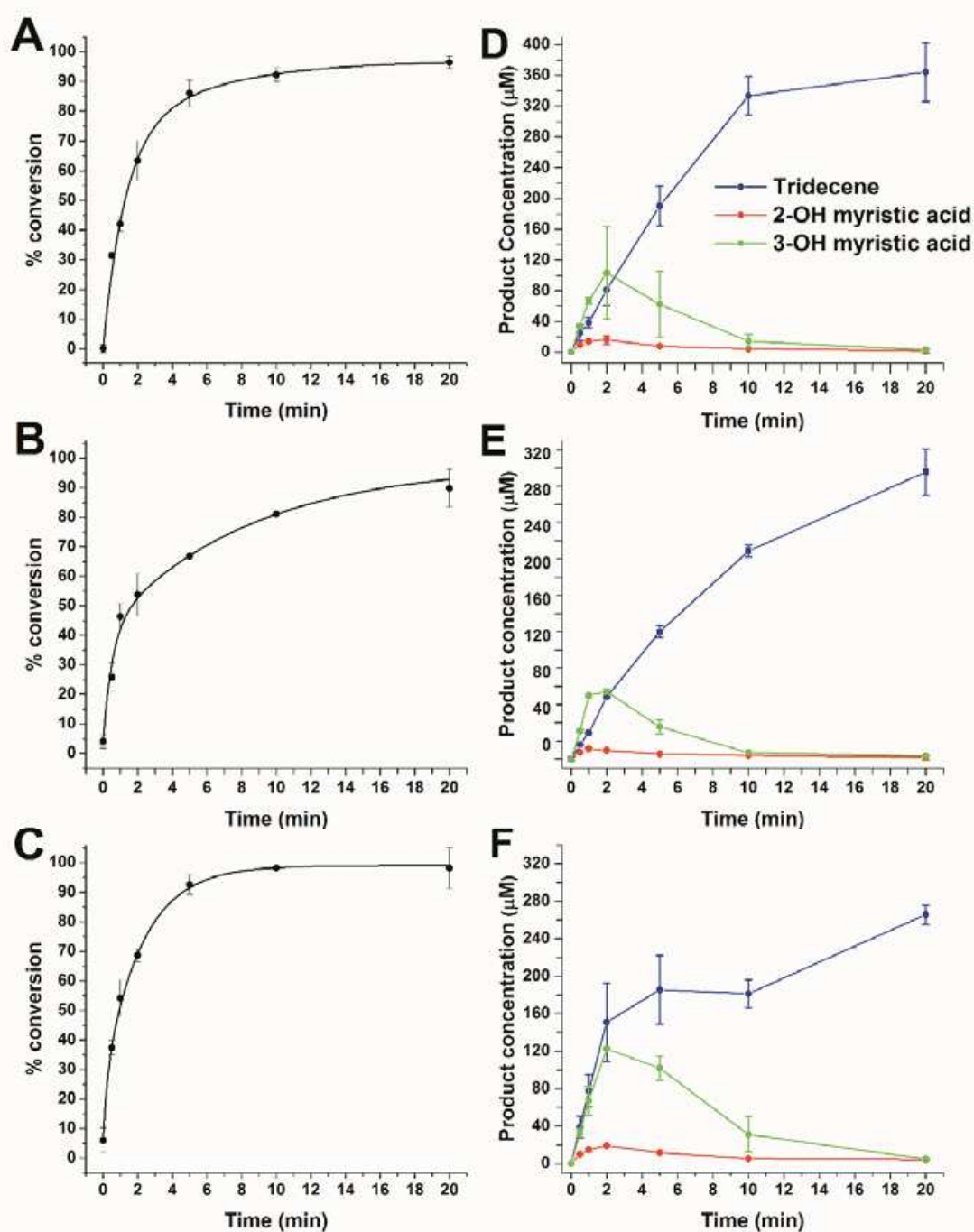


Figure 6



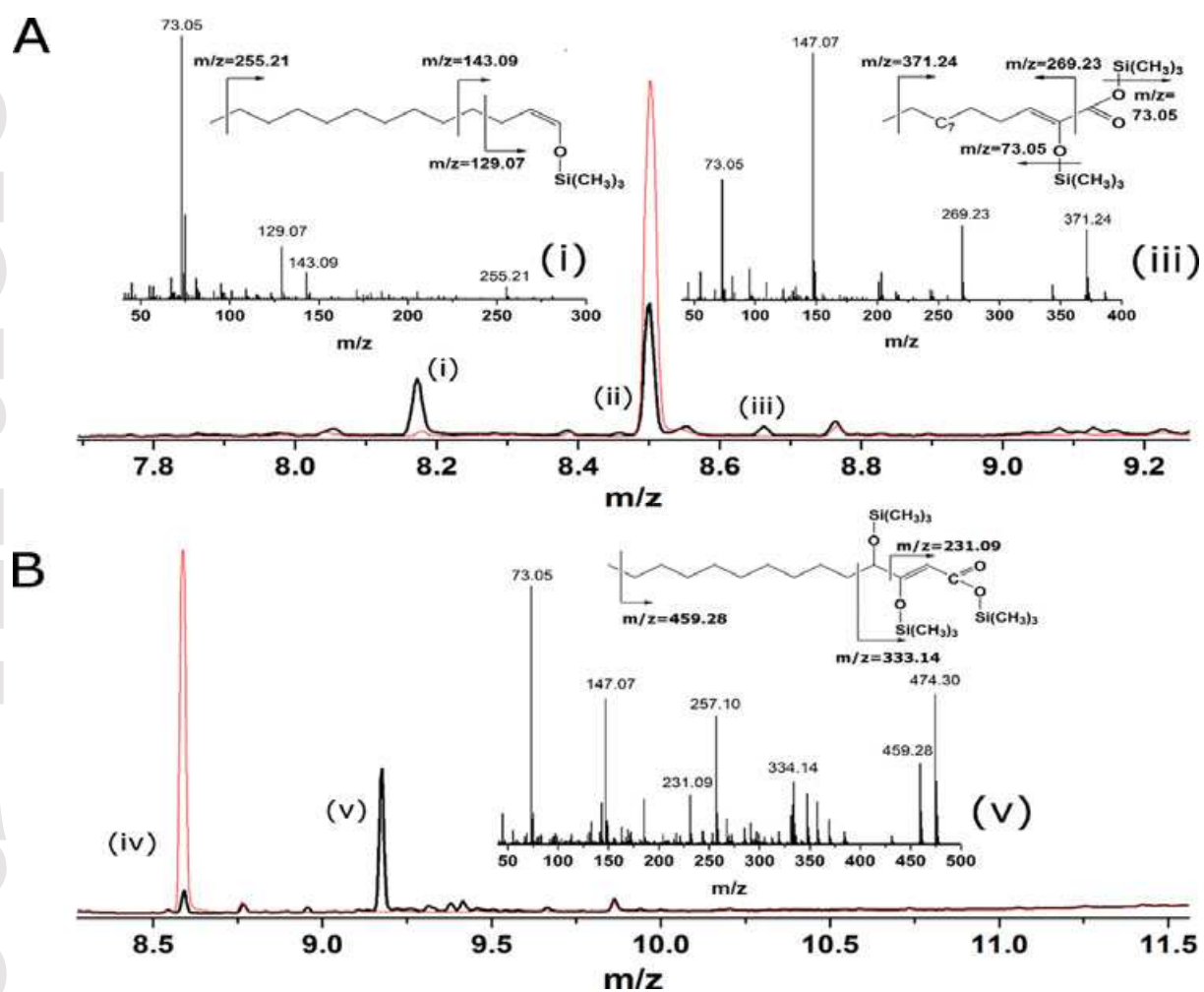


Figure 7

**Table 1. Rates of oxidative modification of OleT<sub>JE</sub>, P450 BM3 heme domain, CYP51B1 and CYP121A1 by H<sub>2</sub>O<sub>2</sub>.** Rate constants for heme oxidation (*k*) and associated absorbance amplitudes (*A*) for heme Soret peak absorption change are presented for OleT<sub>JE</sub> (418 nm), BM3 heme domain (419 nm), CYP121A1 (417 nm) and CYP51B1 (418 nm) across a range of H<sub>2</sub>O<sub>2</sub> concentrations. Data were fitted using a single exponential function. ND indicates that rate constants could not be determined accurately due to small amplitudes of heme absorbance change at the lowest concentrations of H<sub>2</sub>O<sub>2</sub> or (in the case of CYP51B1 and H<sub>2</sub>O<sub>2</sub> concentrations of 0.125 and 0.075 mM) due to linearity of the progress curve.

| H <sub>2</sub> O <sub>2</sub><br>(mM) | OleT <sub>JE</sub>            |               | BM3                           |               | CYP51B1                       |               | CYP121A1                      |               |
|---------------------------------------|-------------------------------|---------------|-------------------------------|---------------|-------------------------------|---------------|-------------------------------|---------------|
|                                       | <i>k</i> (min <sup>-1</sup> ) | <i>A</i>      | <i>k</i> (min <sup>-1</sup> ) | <i>A</i>      | <i>k</i> (min <sup>-1</sup> ) | <i>A</i>      | <i>k</i> (min <sup>-1</sup> ) | <i>A</i>      |
| 2                                     | 8.51 ± 0.17                   | 0.182 ± 0.001 | 43.25 ± 0.26                  | 0.315 ± 0.002 | 16.14 ± 0.23                  | 0.269 ± 0.004 | 26.94 ± 0.23                  | 0.221 ± 0.002 |
| 1                                     | 6.99 ± 0.16                   | 0.065 ± 0.001 | 22.15 ± 0.17                  | 0.284 ± 0.002 | 13.84 ± 0.15                  | 0.236 ± 0.009 | 16.29 ± 0.18                  | 0.193 ± 0.001 |
| 0.5                                   | 5.65 ± 0.14                   | 0.043 ± 0.001 | 17.36 ± 0.11                  | 0.257 ± 0.002 | 10.07 ± 0.11                  | 0.131 ± 0.007 | 14.36 ± 0.16                  | 0.203 ± 0.001 |
| 0.25                                  | 4.96 ± 0.13                   | 0.029 ± 0.001 | 12.60 ± 0.16                  | 0.199 ± 0.002 | 6.79 ± 0.21                   | 0.058 ± 0.013 | 9.38 ± 0.11                   | 0.138 ± 0.001 |
| 0.125                                 | 5.38 ± 0.14                   | 0.021 ± 0.001 | 9.08 ± 0.11                   | 0.103 ± 0.001 | ND                            | ND            | 6.96 ± 0.08                   | 0.077 ± 0.001 |
| 0.075                                 | 5.94 ± 0.12                   | 0.018 ± 0.001 | 7.23 ± 0.08                   | 0.062 ± 0.001 | ND                            | ND            | 5.34 ± 0.08                   | 0.023 ± 0.001 |
| 0.05                                  | ND                            | ND            | 4.36 ± 0.06                   | 0.040 ± 0.001 | ND                            | ND            | 2.16 ± 0.05                   | 0.013 ± 0.001 |
| 0.25                                  | ND                            | ND            | ND                            | ND            | ND                            | ND            | ND                            | ND            |

**Table 2. OleT<sub>JE</sub>-dependent oxidation of myristic acid with different substrates.** The percentage conversion of 0.5 mM myristic acid by the OleT<sub>JE</sub>-HRV3C-AldO fusion enzyme is shown using H<sub>2</sub>O<sub>2</sub> and various AldO substrates at different concentrations. A control reaction using 40 units of catalase, myristic acid and OleT<sub>JE</sub>-HRV3C-AldO confirmed that myristic acid oxidation did not occur in the absence of H<sub>2</sub>O<sub>2</sub>. Myristic acid product analysis was done using GC/MS.

| Substrate                            | Fatty acid conversion (%) |
|--------------------------------------|---------------------------|
| 0.5 mM H <sub>2</sub> O <sub>2</sub> | 87 ± 3                    |
| 1% glycerol                          | 95 ± 3                    |
| 3% glycerol                          | 94 ± 4                    |
| 2 mM sorbitol                        | 93 ± 3                    |
| 10 mM sorbitol                       | 93 ± 2                    |
| 2 mM xylitol                         | 91 ± 5                    |
| 10 mM xylitol                        | 92 ± 3                    |

**Table 3. Myristic acid conversion by OleT<sub>JE</sub> fusion enzymes and products formed.** Reactions were performed with 0.5 mM myristic acid using (i) intact OleT<sub>JE</sub>-HRV3C-AldO, (ii) cleaved OleT<sub>JE</sub>-HRV3C-AldO, and (iii) OleT<sub>JE</sub>- $\alpha$ helix-AldO at 0.1% and 1% glycerol concentrations. Reactions were stopped after 20 minutes.

| Enzyme system                                  | Glycerol (%) | Tridecene ( $\mu$ M) | 2-OH myristic acid ( $\mu$ M) | 3-OH myristic acid ( $\mu$ M) | % substrate conversion |
|--|--------------|----------------------|-------------------------------|-------------------------------|------------------------|
| (i) OleT <sub>JE</sub> -HRV3C-AldO             | 1            | 183 $\pm$ 17         | 0.98 $\pm$ 1.40               | 1.6 $\pm$ 0.1                 | 96 $\pm$ 1             |
|  | 0.1          | 175 $\pm$ 6          | 21 $\pm$ 1                    | 93 $\pm$ 3.3                  | 81 $\pm$ 5             |
| (ii) OleT <sub>JE</sub> + AldO                 | 1            | 137 $\pm$ 38         | 1.0 $\pm$ 1.4                 | 1.7 $\pm$ 0.1                 | 88 $\pm$ 3             |
|  | 0.1          | 209 $\pm$ 19         | 22 $\pm$ 2                    | 113 $\pm$ 7                   | 97 $\pm$ 1             |
| (iii) OleT <sub>JE</sub> - $\alpha$ helix-AldO | 1            | 120 $\pm$ 2          | 2.1 $\pm$ 0.2                 | 2.4 $\pm$ 0.5                 | 98 $\pm$ 3             |
|  | 0.1          | 118 $\pm$ 3          | 27 $\pm$ 2                    | 123 $\pm$ 6                   | 93 $\pm$ 2             |